ABSTRACT

Although atopic dermatitis (AD) is characterized by cytokine production predominantly mediated by T helper (Th) 2 cells, AD pathogenesis also involves innate immune and Th1 cells. To optimize the cytokine milieu required for accurate reproduction of AD-related gene expression profile in vitro, we evaluated the expression pattern of CCL22, CCL17, IL5, IL13, IL33, IL25, TSLP, FLG, and LOR in human lesional AD skin and cytokine-stimulated HaCaT cells. An increase in Th2 mediators (IL5, IL13, CCL22, CCL17, IL25, IL33, and TSLP) and a decrease in genes related to cornified cell envelope (filaggrin and loricrin) were observed in human AD lesions. Innate (tumor necrosis factor-α) and/or Th1/Th2 adaptive cytokines (interferon-γ/IL-4) were required for inducing these inflammatory changes in HaCaT cells, implying that a complex network of innate, Th1, and Th2 cytokines drives AD-like changes. Therefore, stimulation with various combinations of cytokines, beyond Th2 polarization, is necessary when HaCaT cell line is used to study genetic changes implicated in AD pathogenesis.

Keywords: Atopic dermatitis; Cytokine; In vitro stimulation

INTRODUCTION

Atopic dermatitis (AD) has been considered as a T helper (Th) cell type 2 disease characterized by predominant Th2-mediated cytokine production, including IL-4, IL-5, and IL-13, elevated serum IgE, and eosinophilia (1,2). Thymic stromal lymphopoietin (TSLP), IL-25, and IL-33 are mainly produced by epithelial cells, and have important functions of inducing Th2-type adaptive responses and group 2 innate lymphoid cells, which contribute to AD phenotype (3,4). However, Th1 cells and innate inflammatory cytokines are also involved in the pathogenesis of AD. Interferon-γ (IFN-γ) and IL-12 are also expressed in chronic AD lesions. The expression of IFN-γ, but not IL-4, in skin correlates with the clinical course of AD and is known to be downregulated with AD improvement (5). AD patients have elevated levels of CC chemokine receptor 4 ligands, including thymus and activation-regulated chemokine (TARC) and macrophage-derived chemokine (MDC), which mediate preferential Th2 recruitment (6). Additionally, CXC chemokine receptor 3 ligands, which induce Th1
polarization, such as monokine induced by IFN-γ, are also elevated in AD patients compared to normal controls (6). In addition to inflammatory microenvironment imbalance, important barrier-related proteins including filaggrin (FLG) and loricrin (LOR) are decreased in AD (7). In primary keratinocytes, tumor necrosis factor (TNF)-α downregulates FLG and LOR through c-Jun N-terminal kinase (8). These results suggest that complex inflammatory networks, involving innate, Th1, and Th2 responses, orchestrate AD pathogenesis. The aim of this study was to investigate the effects of innate, Th1, and Th2 inflammatory cytokines on the expression of genes implicated in the pathogenesis of AD using human keratinocyte cell line HaCaT.

MATERIALS AND METHODS

HaCaT cell culture and cytokine stimulation
HaCaT cells were grown to 100% confluence and starved for 24 hours in DMEM without fetal bovine serum, followed by their stimulation with various combinations of innate (TNF-α, 10 ng/ml; PeptroTech, Rocky Hill, NJ, USA), Th1 (IFN-γ, 10 ng/ml; PeptroTech), and Th2 (IL-4, 50 ng/ml; PeptroTech) cytokines for 24 hours (9-11). The morphological changes in cells were examined using a phase-contrast microscope (Olympus CX41; Olympus, Tokyo, Japan).

Cell viability assay
The effects of cytokine stimulation on HaCaT cell growth were determined using a water-soluble tetrazolium salt assay kit according to the manufacturer’s instructions (EZ-Cytox cell viability assay kit; ITSBio, Seoul, Korea).

Human sample
Skin specimens from six patients with AD and 12 healthy controls were collected under the approval of the Institutional Review Board at Gachon University Gil Medical Center (GBIRB2016-082). All 6 AD patients received no treatment for at least 4 weeks and had chronic disease (average duration 6.83±4.40 years) with a recent exacerbation in 1–3 months (Table 1).

Real-time PCR validation
Total RNA from skin specimens and HaCaT cells was extracted using the RNeasy Mini Kit (Qiagen, Hilden, Germany). The viable HaCaT cells were harvested with density centrifugation using 40% Percoll (Sigma-Aldrich, St. Louis, MO, USA) and used for RNA extraction. The mRNA level of CCL22 (encoding MDC), CCL17 (encoding TARC), IL5, IL13, FLG, LOR, IL25, TSLP, and IL33 in cytokine-stimulated HaCaT cells was evaluated by real-time PCR and compared with the expression profiles examined in the lesional skin of AD patients. Primer sequences are listed in Table 2.

| Table 1. Characteristics of patients with atopic dermatitis |
|----------------|----------------|----------------|----------------|----------------|
| Patient No. | Sex/age | Duration (yr) | Aggravation (mon) | Co-morbidities |
| 1          | M/20   | 10             | 3               | Asthma         |
| 2          | M/19   | 12             | 2               | Allergic rhinitis |
| 3          | M/21   | 2              | 2               | Allergic rhinitis |
| 4          | M/13   | 5              | 2               | Urticaria      |
| 5          | M/25   | 10             | 1               | -              |
| 6          | M/46   | 2              | 3               | -              |

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**RESULTS**

The viability of HaCaT cells was unaltered following treatment with TNF-α, IFN-γ, or IL-4 alone or the combination of TNF-α and IL-4 (Fig. 1). Moreover, combinatorial treatment of IFN-γ and IL-4 did not affect the viability of HaCaT cells (data now shown). However, treatment of HaCaT cells with the combination of TNF-α and IFN-γ resulted in a significant reduction in the cell viability (Fig. 1). This decrease in the viability of cells was likely attributable to the synergistic cytotoxic effects of IFN-γ and TNF-α. Next, we compared the gene expression profiles of HaCaT cells stimulated with various combinations of cytokines with those observed in human AD skin samples (Fig. 2). Consistent with previous reports (1,2,5,7), the expression of genes encoding Th2 chemokines (CCL22 and CCL17) and Th2 cytokines (IL5, IL13, IL25, and TSLP) was increased in human AD skin lesions, while the expression of genes related to the cornified cell envelope (FLG and LOR) was reduced. The expression of IL-33, a recently described Th2-linked cytokine (12), was significantly decreased in AD skin samples. The gene expression profiles observed for HaCaT cells stimulated with TNF-α and/or IFN-γ were similar to those observed in human AD skin samples. Treatment with IFN-γ alone or TNF-α and IFN-γ combination augmented the expression of CCL22 and CCL17 and decreased the expression of FLG and LOR. Expression of IL-25 and TSLP was increased by combined stimulation with IL-4 and TNF-α. IL-4 treatment inhibited the expression of CCL22 and CCL17 induced by TNF-α and IFN-γ in HaCaT cells, as previously reported (13,14). Furthermore, FLG expression increased upon treatment of HaCaT cells with IL-4 alone but decreased in the presence of the combination of IL-4 and TNF-α. These results imply that the innate and Th1 inflammatory cytokines are required to reproduce AD-like features in HaCaT cells. Stimulation of HaCaT cells with TNF-α and IFN-γ significantly upregulated the expression of IL-33, a Th2-linked cytokine (12).
of *IL33* gene. The expression of classical Th2 cytokines IL-5, IL-13, and IL-25 was markedly upregulated in HaCaT cells following treatment with IL-4.

**DISCUSSION**

AD is a complex inflammatory skin condition, which is incompletely understood. Epidermal barrier defects and dysregulated Th2 immune responses have crucial roles in the pathogenesis of AD. However, it has been reported that human keratinocytes are more sensitive to Th1-
derived lymphocytes than those derived from Th2 in terms of chemokine release (15). In addition, IL-4 influences Th1-type responses, including antigen-induced arthritis, autoimmune uveoretinitis, and T cell transfer model of colitis (16-18). TNF-α along with Th2 cytokines plays an important role to induce AD-like features in epidermal differentiation proteins (19). Our observations have indicated that innate and Th1-type cytokines induce pathogenic changes implicated in the development of AD in HaCaT cells. Although primary human epidermal keratinocytes could be a primary choice to study molecular characteristics of skin, immortalized keratinocytes such as HaCaT cells may be an alternative model, considering their usefulness for in vitro assays (20). HaCaT cells stimulated with cytokines successfully reproduced inflammatory changes, except for IL-33 expression, observed in skin lesions of AD patients (Fig. 2). IL-33, IL-25, and TSLP are known to drive type 2 innate lymphoid cell response (3,4). Despite their similarities, IL-25 and IL-33 have distinct features. IL-25 is constitutively expressed in cellular compartments of epithelial cells and released upon exposure to allergen proteases, whereas IL-33 is a nuclear protein, which has regulatory proteins in its nuclear localization (21), and acts as an alarming signal once it is released by damaged epithelial cells (22). Because the number of mast cells is also increased in AD, overproduction of mast cell chymase could degrade IL-33 and affect its balance in AD (23). In addition, IFN-γ is another key molecule which is reported to upregulate IL-33 levels in keratinocytes of AD patients (9,24). Since most AD patients in this study exhibited chronic disease with recent exacerbation.
and decreased IFNG expression (data not shown), it is plausible that disease status affects the cytokine expression profiles in the lesion of AD. Therefore, cytokine-stimulated HaCaT cell line could be an appropriate tool for the demonstration of mixed chronic and aggravated status of AD. This hypothesis is further supported by the remarkable decrease in cell viability of HaCaT cells upon stimulation with TNF-α and IFN-γ (Fig. 1) and increased expression of IL-33 from damaged HaCaT cells. Taken together, our data suggest that Th1 and Th2 cytokines do not function dichotomously and that a complicated inflammatory network drives AD-like changes. Therefore, future in vitro experiments using HaCaT cells should employ various cytokine combinations beyond Th2 polarization to observe optimal expression of AD-related genes.

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